

2 hr), left in the refrigerator overnight, filtered and solvent removed. A small portion of the Et<sub>2</sub>O-soluble fraction when submitted to TLC [*n*-hexane-Et<sub>2</sub>O (2:1), 2 developments] gave **1** which was purified by further TLC (same solvent system). Another portion of the Et<sub>2</sub>O-soluble fraction when submitted to Silica gel CC (120 g packed in *n*-hexane) gave nine fractions: 1 [*n*-hexane-Et<sub>2</sub>O (2:1)], 2-4 [*n*-hexane-Et<sub>2</sub>O (1:1)], 5-8 (Et<sub>2</sub>O) and 9 [CH<sub>2</sub>Cl<sub>2</sub>-MeOH (1:1)], from which 2-4 were isolated as follows.

Fraction I, which contained **2**, was initially submitted to Silica gel CC and the fraction eluted with *n*-hexane-Et<sub>2</sub>O (4:1) gave **2** on further TLC [*n*-hexane-Et<sub>2</sub>O-AcOH (25:10:1)].

Fractions 6-7 gave **3** when submitted to TLC [*n*-hexane-CHCl<sub>3</sub>-AcOH (10:10:3), 1 development].

TLC of Fr. 2 [*n*-hexane-Et<sub>2</sub>O-AcOH (15:9:1), 2 developments] gave **4**, <sup>13</sup>C NMR (δ, CDCl<sub>3</sub>) (for C-1 to C-20, in order): 27.1, 64.1, 135.8, 144.4, 37.9 or 38.3, 35.2, 27.1, 36.0, 37.9 or 38.3, 41.1, 38.6, 17.4, 125.6, 142.4, 111.0, 138.3, 172.2, 18.6 or 18.8, 15.7 and 18.6 or 18.8, in accord with the structure.

The spectral properties of **1-4** are described in the text.

**Acknowledgments**—We thank Dr S. P. McLaughlin for plant collection and identification and Mr Peter Baker for mass spectral data. This work was supported by the National Science Foundation (Grant PCM-8304771).

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## A NEW CLERODANE DERIVATIVE FROM *TINOSPORA CORDIFOLIA*

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**Key Word Index**—*Tinospora cordifolia*, Menispermaceae, new clerodane diterpenoid.

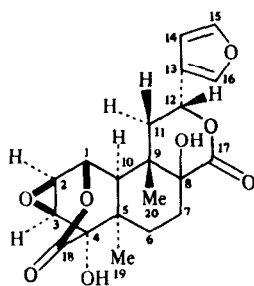
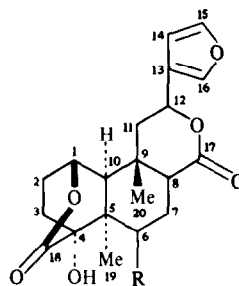
**Abstract**—A new clerodane diterpenoid has been isolated from the stems of *Tinospora cordifolia*. Its structure was established by spectroscopic means and by comparison with closely related clerodane derivatives.

### INTRODUCTION

Chemical investigation of *Tinospora cordifolia* has led to the isolation of a phenolic lignan and one diterpenic furano lactone which were reported earlier [1, 2]. In the

present paper the isolation and structure of a new clerodane derivative, isolated from the same plant, is reported. The structure (**1**) was deduced mainly with the help of <sup>1</sup>H NMR, spin-decoupling and <sup>13</sup>C NMR studies along with the comparison of the spectral data of closely related clerodane diterpenoids such as 6-hydroxy arcangelisin (**2**) [3], fibraurin (**3a**) [4], 6-hydroxy fibraurin (**3b**) [3, 4], palmarin (**4**) [5-7] and 8-hydroxy columbin (**5**) [8].

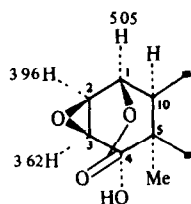
\* Author to whom correspondence should be addressed.

**1****2** R = OH, 2,3 - epoxide**3a** R = H, 2,3 - epoxide, 7,8 - dehydro**3b** R = OH, 2,3 - epoxide, 7,8 - dehydro**4** R = H, 2,3 - epoxide**5** R = H, 8 hydroxy, 2,3 - dehydro

## RESULTS AND DISCUSSION

The molecular formula  $C_{20}H_{22}O_8$  for the terpene was established from its mass spectrum  $[M]^+$ ,  $m/z$  390 and elemental analysis. The IR spectrum (Table 1) showed characteristic absorption for hydroxyl groups ( $3500$ ,  $3480\text{ cm}^{-1}$ ), two lactone carbonyls ( $1750$ ,  $1720\text{ cm}^{-1}$ ), a furan ring ( $3180$ ,  $1510$ ,  $1020$ ,  $880\text{ cm}^{-1}$ , positive Ehrlich test) and an epoxide ring ( $3160$ ,  $1210\text{ cm}^{-1}$  symmetrical stretchings,  $950$ – $850$  and  $860$ – $740\text{ cm}^{-1}$  asymmetrical stretchings)

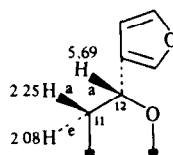
The  $^1\text{H}$ NMR spectrum of the new compound was very similar to that of other furanoditerpenoids and the assignments are given in the Table 2. The signals at  $\delta$  7.71 (1H, *br s*),  $\delta$  7.67 (1H, *br s*) and  $\delta$  6.55 (1H, *br s*) were assigned to two  $\alpha$ - and one  $\beta$ -protons of a  $\beta$ -substituted furan moiety. Two angular methyl groups were observed as singlets at  $\delta$  1.06 and 1.11 (3H each). Two  $\text{D}_2\text{O}$  exchangeable singlets at  $\delta$  6.29 and 6.08 (1H each) were observed and assigned to two tertiary hydroxyl groups. The signals at  $\delta$  5.69 (1H, *dd*,  $J_1 = 11.8$ ,  $J_2 = 5.1\text{ Hz}$ ),  $\delta$  2.25 (1H, *dd*,  $J_1 = 14.3$ ,  $J_2 = 12.0\text{ Hz}$ ) and  $\delta$  2.08 (1H,



$$J_{1,2} = 1.9\text{ Hz}$$

$$J_{2,3} = 4.1\text{ Hz}$$

A

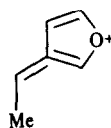


$$J_{11a,12a} = 1.2\text{ Hz}$$

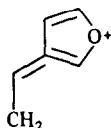
$$J_{11e,12a} = 5.1\text{ Hz}$$

$$J_{11a,11e} = 14.3\text{ Hz}$$

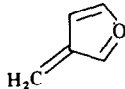
B



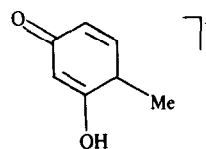
a



b



c



d

## Short Reports

Table 1 Infrared spectral data of diterpenoid furanolactones

Functional groups	1	2	3a	3b	4	5
OH	3500 3480	3300	3460	3500 3400	3470	3500 3450 3400
$\delta$ -Lactones	1750 1720	1780 1705	1766 1690	1773 1717	1771 1700	1750 1735 1705
Furan ring	3180 1510 1020 880 800	1508 1022 875 812	3120 1600 1505 1021	1507 873 810 874 815	1504 881	
Epoxide	3160 1210 950–850 860–740				3150	

\* Absorption frequencies are in  $\text{cm}^{-1}$ , recorded in NujolTable 2  $^1\text{H}$  NMR spectral data of diterpenoid furanolactones

H	1*	2	3a	3b	4	5
1	5.05 ( <i>d</i> , $J = 1.9$ )	5.00	5.08	5.10	4.93	5.54
2	3.96 ( <i>dd</i> , $J_1 = 4.1$ , $J_2 = 1.9$ )	3.88	3.87	3.91	3.80	6.68
3	3.62 ( <i>d</i> , $J = 4.1$ )	3.67	3.65	3.74	3.60	6.42
6	1.60 ( <i>dd</i> , $J_1 = 14.8$ , $J_2 = 8.9$ )	-	1.71 and	4.33		1.50- 1.65
6	1.51 ( <i>dd</i> , $J_1 = 14.4$ , $J_2 = 8.9$ )	3.92	2.30			
7	2.85 ( <i>m</i> )	2.07				3.06
7	1.34 ( <i>m</i> )	1.65	7.25	6.86		1.50- 1.65
8	—	2.97	-		2.74	-
10	1.95 ( <i>br s</i> )	1.78	1.74	1.90	1.60	1.86
11	2.25 ( <i>dd</i> , $J_1 = 14.3$ , $J_2 = 12.0$ )	1.88 and	1.97 and	2.04 and		2.25
11	2.08 ( <i>dd</i> , $J_1 = 14.3$ , $J_2 = 5.1$ )	2.31	2.30	2.29		2.40
12	5.69 ( <i>dd</i> , $J_1 = 11.8$ , $J_2 = 5.1$ )	5.46	5.60	5.68	5.46	5.76
14	6.55 ( <i>br s</i> )	6.65	6.64	6.64	6.63	6.69
15	7.67 ( <i>br s</i> )	7.68	7.67	7.69	7.62	7.63
16	7.71 ( <i>br s</i> )	7.75	7.76	7.76	7.70	7.77
19	1.11 ( <i>q</i> )	1.26	1.06 1.15	1.16	1.18	1.20
20	1.06 ( <i>q</i> )	1.05		1.18	1.04	1.00
4-OH	6.29 ( <i>br s</i> )	6.06	6.38	6.34	6.24	5.16
6-OH	—	5.54	-	5.51		
8-OH	6.08 ( <i>br s</i> )	—	-	-	-	4.83

\* At 500 MHz, chemical shifts are in  $\delta$  values from TMS, coupling constants ( $J$ ) in Hz, in DMSO- $d_6$  solution

$dd$ ,  $J_1 = 14.3$ ,  $J_2 = 5.1$  Hz) were assigned to an 'ABX' system as given in the part structure 'B' (C-11, C-12). Two multiplets were observed at  $\delta 2.85$  and  $1.34$  (1H each) which were assigned to protons at C-7. The down field signal at  $\delta 2.85$  was assigned to an axial proton on C-7 which experiences the anisotropic effect of the lactone carbonyl group. The signals at  $\delta 5.05$  (1H,  $d$ ,  $J = 1.9$  Hz),  $\delta 3.96$  (1H,  $dd$ ,  $J_1 = 4.1$ ,  $J_2 = 1.9$  Hz) and  $\delta 3.62$  (1H,  $d$ ,  $J = 4.1$  Hz) could be explained by considering an epoxide ring in the part structure 'A'. The presence of partial structures 'A' and 'B' were confirmed on the basis of 100 MHz  $^1\text{H}$  NMR spin-decoupling experiments.

On irradiating the signal at  $\delta 5.05$  (H-1), the signal at  $\delta 3.96$  (H-2) was collapsed into a doublet ( $J_{2,3} = 4.1$  Hz). On irradiating the signal at  $\delta 3.96$  (H-2), the signals at  $\delta 5.05$  (H-1) and  $\delta 3.62$  (H-3) were collapsed into two clean singlets. These observations supported the presence of partial structure 'A'.

When the signal at  $\delta 5.69$  (H-12, X of ABX) was irradiated, the signals at  $\delta 2.25$  and  $2.08$  ( $H_a$ -11 and  $H_e$ -11, AB of ABX) collapsed into two doublets ( $J_{AB} = 14.3$  Hz). Irradiation of the signal at  $\delta 2.25$  ( $H_a$ -11, B of ABX), caused the signals at  $\delta 5.69$  and  $2.08$  ( $H_a$ -12 and  $H_e$ -11, AX of ABX system) to collapse into two doublets ( $J_{AX} = 5.1$  Hz). These observations suggested a partial structure 'B' having an axial proton at C-12 and vicinal methylene protons at C-11. A broad singlet was observed for H-10 showing a very small coupling constant between the vicinal axial-equatorial protons (H-1, H-10) [9]. This clearly indicated the *trans* diaxial relationship of the C-1 lactone oxygen and H-10.

The stereochemistry of the epoxide is deduced from the coupling constant values between H-1, H-2 and between H-2, H-3. For  $\beta$ -epoxides these values are of the order of 2-3 Hz and 4.2 Hz, respectively (Table 3). In compound

Table 3 Coupling constants between H-1, H-2 and H-2, H-3 in the 2,3- $\beta$  epoxide moiety of diterpenoid furanolactones

	$J_{1,2}^*$	$J_{2,3}^*$	Ref
Fibraurin	2.9	4.2	[4]
A derivative of fibraurin	2.2	4.2	[4]
Palmarin	3.0	4.2	[7]
A derivative of palmarin	2.0	4.0	[7]
Compound (1)	1.9	4.1	—

\*Coupling constants ( $J$ ) in Hz

(1) the coupling constants between H-1, H-2 and H-2, H-3 are 1.9 and 4.1 Hz, respectively. Hence, the epoxide group present in the compound could be  $\beta$ .

The noise decoupled and single frequency off-resonance  $^{13}\text{C}$  NMR spectrum contained signals arising from two methyl carbons ( $q$ ,  $\delta 20.1$  and  $22.8$ ), three methylene carbons ( $t$ ,  $\delta 26.1$ ,  $26.6$  and  $34.5$ ), five methine carbons ( $d$ ,  $\delta 45.6$ ,  $49.2$ ,  $51.1$ ,  $69.9$  and  $70.6$ ), three furanoid carbons ( $d$ ,  $\delta 109.1$ ,  $140.1$  and  $143.9$ ), five quaternary carbons ( $s$ ,  $\delta 39.0$ ,  $40.5$ ,  $71.7$ ,  $80.4$  and  $125.5$ ) and two lactone carbonyl carbons ( $s$ ,  $\delta 171.6$  and  $173.1$ ). These values compared well with the values reported for clerodane derivatives (Table 4).

Manbe and Nishino [10] reported the usefulness of  $^{13}\text{C}$  chemical shifts of angular methyls to distinguish between *cis* and *trans* A/B-ring junction in clerodane diterpenes. In the case of *cis* clerodanes, the C-19 methyl carbon atom resonates in a region above  $\delta 20$  and in corresponding *trans* compounds it resonates in the region  $\delta 11$ – $19$ . The C-19 methyl signal of compound (1) was found at  $\delta 22.8$ , and hence it is concluded that the A/B-ring junction is *cis*.

Table 4  $^{13}\text{C}$  NMR chemical shifts of diterpenoid furanolactones

C	1*	2	3a	3b	4	5
1	69.9 <i>d</i>	69.2 <i>d</i>	69.7 <i>d</i>	69.9 <i>d</i>	69.4 <i>d</i>	73.1 <i>d</i>
2	49.2 <i>d</i>	49.1 <i>d</i>	49.5 <i>d</i>	49.4 <i>d</i>	49.2 <i>d</i>	130.3 <i>s</i>
3	51.1 <i>d</i>	50.5 <i>d</i>	51.6 <i>d</i>	51.1 <i>d</i>	50.9 <i>d</i>	136.5 <i>s</i>
4	80.4 <i>s</i>	82.7 <i>s</i>	80.0 <i>s</i>	82.0 <i>s</i>	80.3 <i>s</i>	80.6 <i>s</i>
5	38.9 <i>s</i>	34.8 <i>s</i>	35.2 <i>s</i>	35.2 <i>s</i>	40.4 <i>s</i>	37.0 <i>s</i>
6	26.1 <i>t</i>	69.3 <i>d</i>	31.2 <i>t</i>	70.2 <i>d</i>	25.3 <i>t</i>	26.6 <i>t</i>
7	26.6 <i>t</i>	27.3 <i>t</i>	142.1 <i>d</i>	145.8 <i>d</i>	16.5 <i>t</i>	26.7 <i>t</i>
8	71.7 <i>s</i>	43.1 <i>d</i>	133.7 <i>s</i>	131.1 <i>s</i>	41.8 <i>d</i>	72.2 <i>s</i>
9	40.5 <i>s</i>	43.4 <i>s</i>	44.8 <i>s</i>	48.7 <i>s</i>	34.6 <i>s</i>	39.5 <i>s</i>
10	45.6 <i>d</i>	55.5 <i>d</i>	54.0 <i>d</i>	55.1 <i>d</i>	53.0 <i>d</i>	46.5 <i>d</i>
11	34.5 <i>t</i>	43.2 <i>t</i>	42.0 <i>t</i>	42.0 <i>t</i>	43.2 <i>t</i>	34.7 <i>t</i>
12	70.6 <i>d</i>	71.3 <i>d</i>	70.6 <i>d</i>	71.0 <i>d</i>	70.9 <i>d</i>	70.6 <i>d</i>
13	125.5 <i>s</i>	125.2 <i>s</i>	125.0 <i>s</i>	124.8 <i>s</i>	125.0 <i>s</i>	125.5 <i>s</i>
14	109.1 <i>d</i>	109.1 <i>d</i>	109.1 <i>d</i>	109.0 <i>d</i>	108.8 <i>d</i>	109.1 <i>d</i>
15	140.1 <i>d</i>	140.4 <i>d</i>	140.4 <i>d</i>	140.4 <i>d</i>	139.9 <i>d</i>	140.2 <i>d</i>
16	143.9 <i>d</i>	143.8 <i>d</i>	143.9 <i>d</i>	143.9 <i>d</i>	143.4 <i>d</i>	143.8 <i>d</i>
17	173.1 <i>s</i>	171.7 <i>s</i>	171.5 <i>s</i>	171.1 <i>s</i>	172.0 <i>s</i>	174.5 <i>s</i>
18	171.6 <i>s</i>	171.4 <i>s</i>	163.2 <i>s</i>	169.1 <i>s</i>	172.9 <i>s</i>	171.2 <i>s</i>
19	22.8 <i>q</i>	17.2 <i>q</i>	25.0 <i>q</i>	21.3 <i>q</i>	23.6 <i>q</i>	24.0 <i>q</i>
20	20.1 <i>q</i>	18.4 <i>q</i>	20.7 <i>q</i>	18.4 <i>q</i>	17.4 <i>q</i>	19.8 <i>q</i>

\*Chemical shifts are in  $\delta$  values from TMS at 125 MHz, in DMSO- $d_6$  solution.

The proposed structure (I) as a diterpenoid furano lactone clearly satisfied the above spectral data. The mass fragmentation pattern also supported the structure (I). The mass spectrum gave peaks at  $m/z$  390  $[M]^+$ , 346  $[M - CO_2]^+$ , 95, 94 and 81 which are due to the fragments 'a', 'b' and 'c', respectively, in accord with similar diterpenoid furano lactones [4, 11]. A characteristic and base peak at  $m/z$  124 is assigned to the ion 'd' which arises by the retro-Diels-Alder type fragmentation of ring B.

#### EXPERIMENTAL

Mps. uncorr. IR spectra were recorded in Nujol. The proton spin-decoupling NMR expts were carried out at 100 MHz.

**Extraction and isolation of (I).** Stems of *T. cordifolia* Miex. (26 kg) were collected from IIT Campus, Bombay and identified by Dr. Agarkar, Institute of Science, Bombay. The stems were dried, finely powdered and extd with  $CHCl_3$  (60 l) in a Soxhlet for 48 hr. Repeated CC over silica gel with 70% EtOAc-petrol (60–80°) (7:3) afforded (I) which was recrystallized from MeOH (60:8 g, 3%:10%:3%) as cubic crystals, mp. 231–233° (decomp.),  $[\alpha]_D^{20} = +28.2$  (DMSO,  $c$  0.62). IR  $\nu_{max}^{nujol}$   $cm^{-1}$  (Table 1), UV  $\lambda_{max}^{MeOH}$   $\log \epsilon$  207.8 nm (3.8).  $^1H$ -NMR (500 MHz, DMSO- $d_6$ ) (Table 2),  $^{13}C$ -NMR (125 MHz, DMSO- $d_6$ ) (Table 4), MS  $m/z$  390  $[M]^+$  (0.5%), 346  $[M - CO_2]^+$  (1), 291 (1), 252 (25), 199 (2), 125 (24), 124 (100), 95 (38), 94 (32), 93 (12), 91 (25), 81 (33). Anal. calcd. for  $C_{20}H_{22}O_8$ : C, 61.53, H, 5.68. Found: C, 61.29, H, 5.54%.

**Acknowledgement**—The use of facilities provided by the 500 MHz FT-NMR National Facility at TIFR, Bombay are gratefully acknowledged.

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## A SOYASAPOGENOL-B GLUCOSIDE FROM THE SEEDS OF *PHASEOLUS VULGARIS*

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**Key Word Index**—*Phaseolus vulgaris* Leguminosae triterpenoid glucoside soyasapogenol B-24-O- $\beta$ -D-glucopyranoside

**Abstract**—A new triterpenoid glucoside has been isolated from the seeds of *Phaseolus vulgaris* and characterized as 3 $\beta$ ,22 $\beta$ -dihydroxy olean-12-en-24-O- $\beta$ -D-glucopyranoside

#### INTRODUCTION

Seeds of *Phaseolus vulgaris* Linn (French bean) are a rich source of saponins [1]. Crude extracts of the seeds show antifertility activity [2] and are known to contain phyto-sterols, triterpenoids and triterpenoid saponins [3–6].

#### RESULTS AND DISCUSSION

The new triterpenoid glucoside (1) was isolated from a methanolic extract of the seeds by column chromatography and purified by droplet counter current chromatography (DCCC).